

## Unveiled chromosomal diversity in the Araneidae (Araneomorphae): the highest diploid number among entelegynes and the first record of the $X_1X_2X_3X_4$ Sex Chromosome System in the family

Lucas Henrique Bonfim Souza<sup>1</sup>, Caroline Correia Costa<sup>1</sup>, Bruno Cansação Silva<sup>1</sup>, Debora Duarte Dutra<sup>1</sup>, Arthur dos Santos Montanholi<sup>1</sup>, Beatriz Oliveira<sup>1</sup>, Susan Roghanian<sup>1</sup>, Larissa Candido Lemos<sup>1</sup>, Henrique Ranieri Covali Pontes<sup>1</sup>, Aline Agatha de Pádua<sup>1</sup>, Rebeca Rocha Sobrinho dos Santos Dias<sup>1</sup>, Antonio Domingos Brescovit<sup>2</sup> and Douglas Araujo<sup>1</sup>: <sup>1</sup>Laboratório de Citotaxonomia e Evolução Cromossômica Animal, Universidade Federal de Mato Grosso do Sul, UFMS, Instituto de Biociências, Cidade Universitária, Caixa Postal 549, CEP 79070-900, Campo Grande, Brazil; E-mail: lucashenriquebs18@gmail.com; <sup>2</sup>Laboratório de Coleções Zoológicas, Instituto Butantan, Av. Vital Brasil, 1500, CEP 05503-900, São Paulo, Brazil.

**Abstract.** The Araneidae is among the most speciose spider families, but there are few karyotype studies (1.9%) and some species-rich clades are without any chromosomal study. Understanding the evolution of chromosome number and Sex Chromosome Systems is made more difficult by many uncertain evolutionary relationships within the family. In this work, the chromosomal analysis of eight araneid species (*Acacesia benigna* Glueck, 1994, *Actinosoma pentacanthum* (Walckenaer, 1841), *Alpaida bicornuta* (Taczanowski, 1878), *Dubiepeira* Levi, 1991 sp., *Gasteracantha cancriformis* (Linnaeus, 1758), *Parawixia bistriata* (Rengger, 1836), *Verrucosa meridionalis* (Keyserling, 1892) and *Verrucosa scapofracta* Lise, Kesster & Silva, 2015), contribute to discussions of some evolutionary scenarios of chromosome evolution. The gonads were submitted to colchicine treatment, hypotonization, slide preparation, and Giemsa staining. The species analyzed showed  $2n\delta = 24$  ( $11\text{II} + X_1X_2$ ), except *Dubiepeira* sp. with  $2n\delta = 41$  ( $19\text{II} + X_1X_2X_3$ ), and both *Verrucosa* species, which presented  $2n\delta = 47$  ( $22\text{II} + X_1X_2X_3$ ) in *V. meridionalis* and  $2n\delta = 50$  ( $23\text{II} + X_1X_2X_3X_4$ ) in *V. scapofracta*. The species analyzed possess all chromosomes with acro/telocentric chromosomal morphology. The  $2n\delta = 24$ ,  $X_1X_2$  found in most species studied here is the most frequent karyotype in the Araneidae. This study presents the first chromosomal data for the diverse clade “Micrathenines”, the highest diploid number among entelegynes ( $2n\delta = 50$ ), and the first record of an  $X_1X_2X_3X_4$  in the Araneidae. The chromosome data suggest a series of fission events in the origin of *Verrucosa* karyotypes, and a close relationship between *Dubiepeira* sp. and *Araneus ventricosus* (L. Koch, 1878). Moreover, *Alpaida bicornuta* can be cytotaxonomically distinguished of other *Alpaida* species karyotyped up to now.

**Keywords:** Sex chromosomes, high diploid numbers, Araneae, spiders, Araneoidea.

<https://doi.org/10.1636/JoA-S-20-071>

The Araneidae is the third most speciose spider family, with 3058 species described up to now (World Spider Catalog 2021). This great diversity makes detailed studies of the clade more difficult (Scharff et al. 2020) and, in this scenario, the use of different tools to analyze the species, such as molecular and cytogenetic data, can be helpful in unveiling the evolutionary relationships (Araujo et al. 2011; Scharff et al. 2020).

In the Araneidae, the diploid number varies from  $2n\delta = 13$  in *Neoscona* Simon, 1864 sp. to  $2n\delta = 49$  in *Araneus ventricosus* (L. Koch, 1878) (Parida & Sharma 1987; Sharma & Parida 1987; Youju et al. 1993). The Sex Chromosome Systems (SCS) found in the family are of the types  $\delta X_0/\delta XX$ ,  $\delta X_1X_2/\delta X_1X_1X_2X_2$ ,  $\delta X_1X_2X_3/\delta X_1X_1X_2X_2X_3X_3$  and  $\delta XY/\delta XX$ . Regarding the chromosomal morphology, all types have been described in the Araneidae. However, most analyzed species possess  $2n\delta = 24$  (81%),  $\delta X_1X_2/\delta X_1X_1X_2X_2$  SCS (89%) and acro/telocentric chromosomal morphology (59%) (Araujo et al. 2021). Unfortunately, only 1.9% of the araneid species were karyotyped and some clades that have genera with high species richness remain chromosomally unknown, as the “Micrathenines” clade (Scharff et al. 2020; Araujo et al. 2021).

The aim of this study was to cytogenetically analyze eight araneid species belonging to clades Eriophorines, Gasteracanthines, Micrathenines and two species with phylogenetic position not established, in order to discuss the chromosome

evolution in araneid spiders and contribute with new karyotype data in conflicting and less studied taxonomic groups.

### METHODS

The specimens of all analyzed species (Table 1) were collected by active night search, with exception of *Actinosoma pentacanthum* (Walckenaer, 1841) that was collected during the day on the surface of aquatic plants. The collections were performed in the states of Mato Grosso do Sul (MS) and São Paulo (SP), Brazil, from May of 2014 to April of 2018 and vouchers are deposited in the Laboratório de Coleções Zoológicas, Instituto Butantan, São Paulo, SP, Brazil (IBSP, curator: A.D. Brescovit).

The chromosome preparations of gonads and embryos were carried out following Araujo et al. (2008). All analyzed cells were photographed employing a Zeiss Axioimager D2 microscope with a monochromatic AxioCam 503 camera, using the ZEN Pro software. Chromosome morphology was determined with the software IMAGEJ (Rasband 1997–2020) and the LEVAN plugin (Sakamoto & Zacaro 2009), according to Levan et al. (1964) and Green & Sessions (1991).

### RESULTS

**Karyotypes with the araneid common diploid number.**—The species *Acacesia benigna*, *Actinosoma pentacanthum*, *Alpaida*

Table 1.—Species, collection site and number of specimens analyzed in this work.

Species	Locality	Specimens analyzed	Vouchers (IBSP)
<i>Acacesia benigna</i> Glueck, 1994	Campo Grande, MS (20°30'36"S/54°36'54"W)	1 ♂	222032
<i>Actinosoma pentacanthum</i> (Walckenaer, 1841)	Corumbá, MS (19°34'37"S/57°00'42"W) and Porto Murtinho, MS (21°49'26"S/57°48'48"W)	9 ♂/12 ♀ & 4 embryos	211335, 212177, 212186, 212193, 221987, 222376, 222379 - 222390, 222397, 222401 and 222403
<i>Alpaida bicornuta</i> (Taczanowski, 1878)	Campo Grande, MS (20°30'36"S/54°36'54"W)	2 ♂	222039 and 222040
<i>Dubiepeira</i> sp.	Campo Grande, MS (20°30'36"S/54°36'54"W)	2 ♂/3 ♀	222034 - 222038
<i>Gasteracantha cancriformis</i> (Linnaeus, 1758)	Campo Grande, MS (20°30'36"S/54°36'54"W)	9 ♀ & 6 embryos	166506, 166533, 166538, 166872, 166880, 166893, 166899, 166901, 166922
<i>Parawixia bistrata</i> (Rengger, 1836)	Dois Irmãos do Buriti, MS (20°31'33.83"S/55°25'11.67"W)	3 ♂/4 ♀	211341, 211343, 211346 - 211350
<i>Verrucosa meridionalis</i> (Keyserling, 1892)	Aquidauana, MS (20°27'03.93"S/55°37'17.02"W)/ Campo Grande, MS (20°30'36"S/54°36'54"W)/ Botucatu, SP (22°50'44.3"S/48°25'32.4"W) and Campo Grande, MS (20°29'19.09"S/54°39'39.06"W)	2 ♂/4 ♀	214414, 214415, 221999, 222412, 242323. 166879
<i>Verrucosa scapofracta</i> Lise, Kesster & Silva, 2015	Iguape, SP (24°42'13.03"S/47°32'49.98"W)	1 ♂	222317

MS = state of Mato Grosso do Sul. SP = state of São Paulo.

*bicornuta* and *Parawixia bistrata* showed in all males analyzed mitotic metaphases with  $2n\delta = 24$  (Figs. 1A–D). Females of *A. pentacanthum*, *P. bistrata* and *Gasteracantha cancriformis* showed  $2n\text{♀} = 26$  (Figs. 1E–G). Although no adult *G. cancriformis* male was collected, two embryos presented  $2n = 24$  (considered males) (Fig. 1H), while four other embryos had the same chromosome number as adult females. The four embryos (3 ♂ and 1 ♀) of *A. pentacanthum* analyzed possess the same diploid number as adults.

The morphology of all chromosomes is acro/telocentric, and chromosomes show small differences in length (Fig. 1), with exception of the largest chromosome pair of *A. bicornuta*, which is almost twice the length of the second longest pair (Fig. 1C).

Spermatocytes I in diplotene of *A. benigna*, *A. pentacanthum*, *A. bicornuta* and *P. bistrata* revealed 11 autosomal bivalents and two positively heteropycnotic sex univalents (Figs. 2A–D). The SCS was confirmed as of the type  $X_1X_2\delta$ /

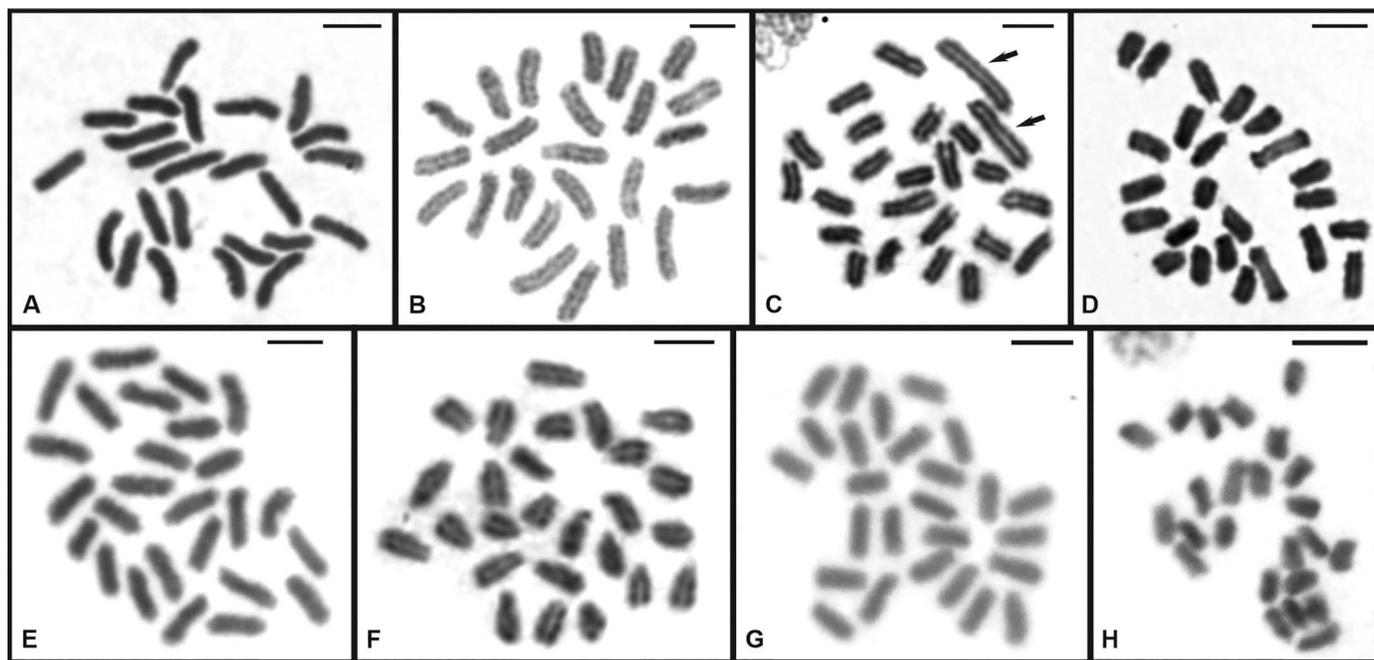


Figure 1A–H.—Araneidae mitotic cells with  $2n\delta = 24$  (A–D, H) and  $2n\text{♀} = 26$  (E–G): A, *Acacesia benigna*. B, E, *Actinosoma pentacanthum*. C, *Alpaida bicornuta*. D, F, *Parawixia bistrata*. G, H, *Gasteracantha cancriformis*. Arrows indicate the largest chromosome pair of *A. bicornuta*. Scale bar: 5  $\mu\text{m}$ .

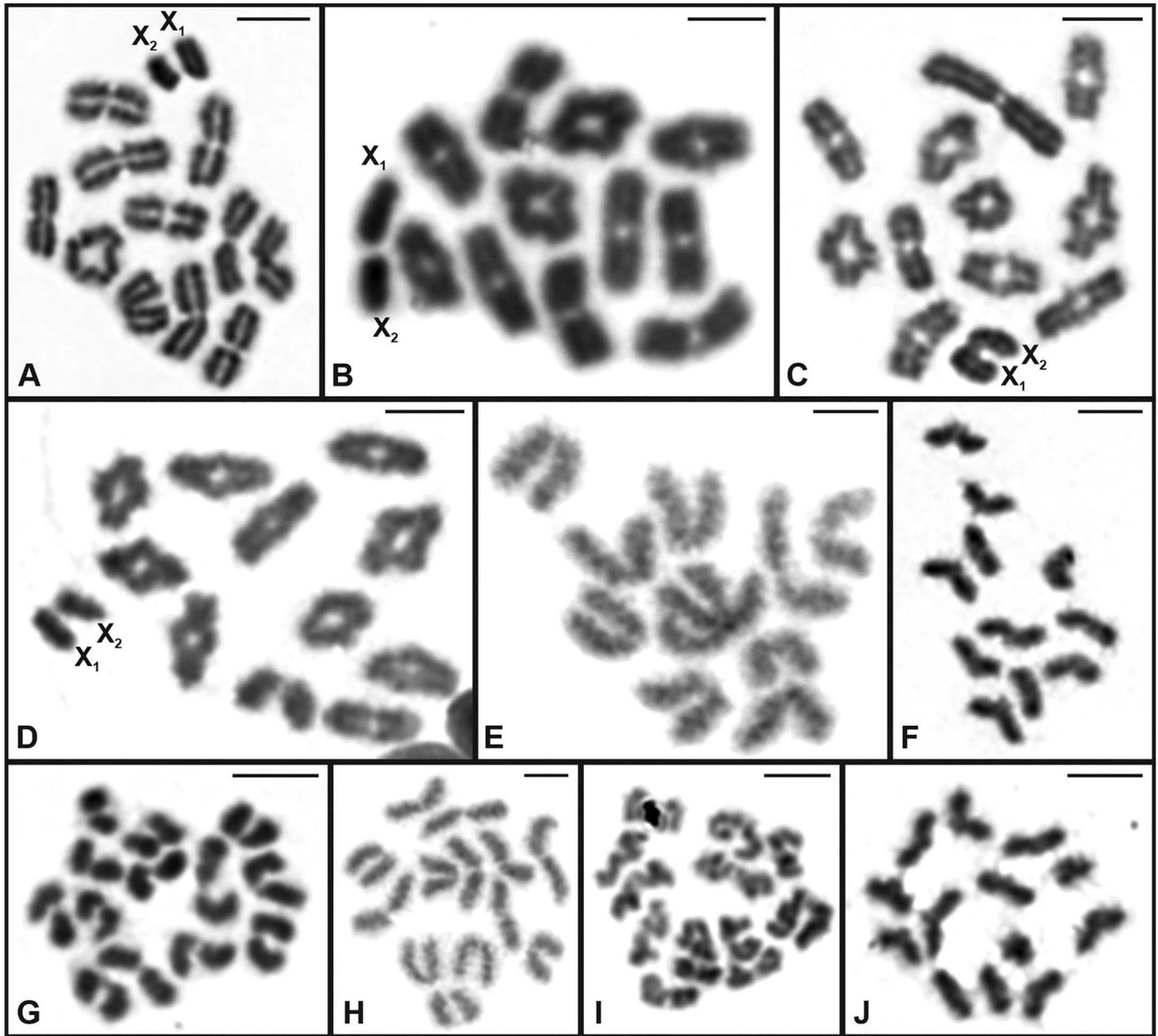


Figure 2A–J.—Meiotic cells of species with the common araneid diploid number ( $2n\delta = 24$ ): A–D. Diplotenes with 11 autosomal bivalents +  $X_1X_2$ . E–J. Metaphase II with  $n = 11$  (E, F) and  $n = 13 = 11 + X_1X_2$  (G–J). A, G. *Acacesia benigna*. B, E, H. *Actinosoma pentacanthum*. C, I. *Alpaida bicornuta*. D, F, J. *Parawixia bistrata*. Scale bar: 5  $\mu$ m.

$X_1X_1X_2X_2\delta$  through spermatocytes II in metaphase with  $n = 11$  in *A. pentacanthum* and *P. bistrata* (Fig. 2E, F) and with  $n = 13 = 11 + X_1X_2$  in these four species (Figs. 2G–J).

Thus, in all the analyzed cells (Table 2) these five species exhibited  $2n\delta = 24 = 22 + X_1X_2$  and  $2n\delta = 26 = 22 + X_1X_1X_2X_2$  and chromosomes with acro/telocentric morphology.

**Karyotypes with high diploid numbers.**—*Dubiepeira* sp. showed males with  $2n\delta = 41$  and females with  $2n\delta = 44$ , with all chromosomes acro/telocentric gradually decreasing in size (Figs. 3A, B). Spermatocytes I in diplotene of *Dubiepeira* sp. exhibit 19 autosomal bivalents and three sex univalents (Fig. 3C). Only spermatocytes II in metaphase with  $n = 19$

were found (Fig. 3D). Altogether, these characteristics confirm the SCS of the type  $X_1X_2X_3\delta/X_1X_1X_2X_2X_3X_3\delta$

*Verrucosa meridionalis* and *V. scapofracta* showed  $2n\delta = 47 / 2n\delta = 50$  and  $2n\delta = 50$ , respectively, with all chromosomes of both species with acro/telocentric chromosome morphology and gradually decreasing in size (Figs. 3E–G). Spermatocytes I in diplotene of *V. meridionalis* possess 22 autosomal bivalents and three positively heteropycnotic sex univalents (Fig. 3H), while the spermatocytes II in metaphase showed  $n = 22$  and  $n = 25 = 22 + X_1X_2X_3$  (Fig. 3I). Spermatocytes I in diplotene from the single male analyzed of *V. scapofracta* exhibit 23 autosomal bivalents and four positively heteropycnotic sex

Table 2.—Number of analyzed cells, diploid number and sex chromosome system (SCS) of species analyzed in this work.

Species	Analyzed cells	Diploid number ( $2n\delta$ )	SCS ( $\delta$ )
<i>Acacesia benigna</i> Glueck, 1994	50	24	$X_1X_2$
<i>Actinosoma pentacanthum</i> (Walckenaer, 1841)	236	24	$X_1X_2$
<i>Alpaida bicornuta</i> Taczanowski, 1878)	87	24	$X_1X_2$
<i>Gasteracantha cancriformis</i> (Linnaeus, 1758)	35	24	$X_1X_2$
<i>Parawixia bistrata</i> (Rengger, 1836) Acro/telocentric	74	24	$X_1X_2$
<i>Dubiepeira</i> sp.	104	41	$X_1X_2X_3$
<i>Verrucosa meridionalis</i> (Keyserling, 1892)	64	47	$X_1X_2X_3$
<i>Verrucosa scapofracta</i> Lise, Kesster & Silva, 2015	10	50	$X_1X_2X_3X_4$

univalent (Fig. 3J), and only spermatocytes II in metaphase with  $n = 27 = 23 + X_1X_2X_3X_4$  were found (Fig. 3K).

Thus, based on all cells analyzed (Table 2) *Dubiepeira* sp. possess  $2n\delta = 41 = 38 + X_1X_2X_3$  and  $2n\varphi = 44 = 38 + X_1X_1X_2X_2X_3X_3$ , *V. meridionalis* has  $2n\delta = 47 = 44 + X_1X_2X_3$

and  $2n\varphi = 50 = 44 + X_1X_1X_2X_2X_3X_3$  and *V. scapofracta*  $2n\delta = 50 = 46 + X_1X_2X_3X_4$  and we suggest that females have the corresponding diploid number  $2n\varphi = 54 = 46 + X_1X_1X_2X_2X_3X_3X_4X_4$ . All species with acro/telocentric chromosomes.

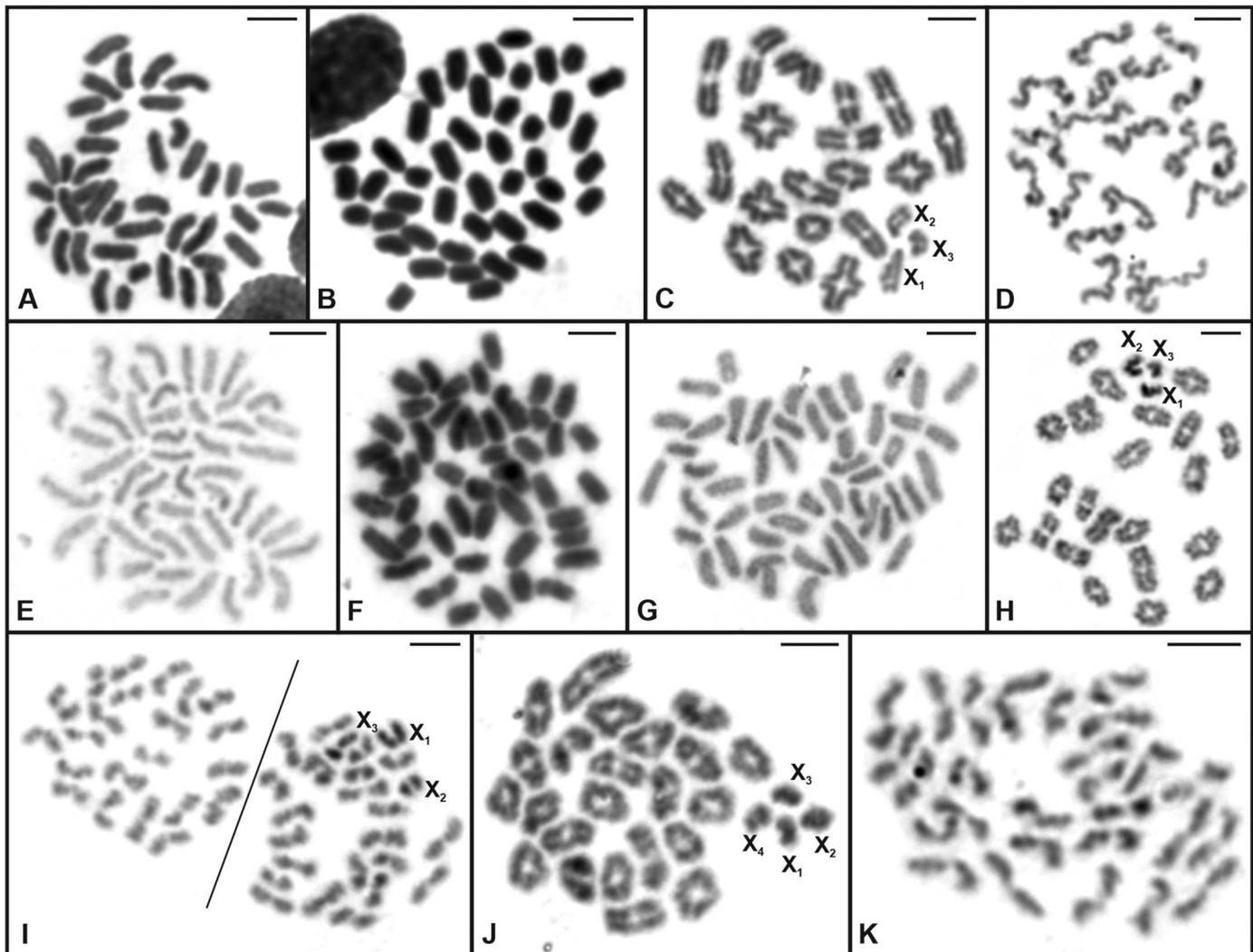


Figure 3A–K.—Cell divisions in *Dubiepeira* sp. (A–D), *Verrucosa meridionalis* (E–F, H–I) and *Verrucosa scapofracta* (G, J–K). A. Spermatogonial metaphase with  $2n\delta = 41$ . B. Oogonial metaphase with  $2n\varphi = 44$ . C. Diplotene with 19 autosomal bivalents +  $X_1X_2X_3$ . D. Metaphase II with  $n = 19$ . E. Spermatogonial metaphase with  $2n\delta = 47$ . F. Oogonial metaphase with  $2n\varphi = 50$ . G. Spermatogonial metaphase with  $2n\delta = 50$ . H. Diplotene with 22 autosomal bivalents +  $X_1X_2X_3$ . I. Metaphase II with  $n = 22$  (left) and  $n = 25 = 22 + X_1X_2X_3$  (right). J. Diplotene with 23 autosomal bivalents +  $X_1X_2X_3X_4$ . K. Metaphase II with  $n = 27 = 23 + X_1X_2X_3X_4$ . Scale bar: 5  $\mu$ m.

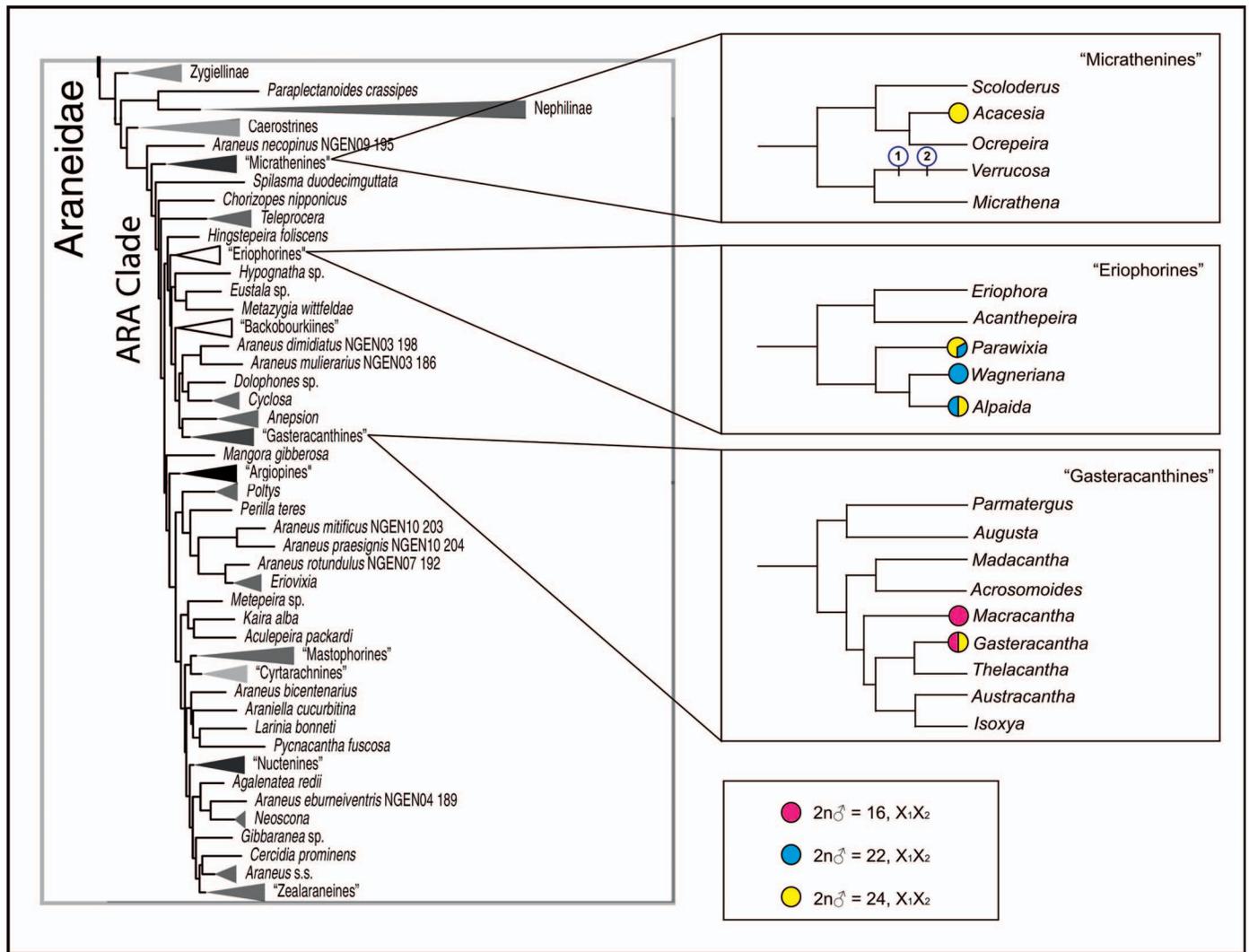


Figure 4.—Araneidae summary tree extracted from Scharff et al. (2020) showing the cytogenetic data available for “Micrathenines,” “Eriophorines,” and “Gasteracanthines” in detail.

DISCUSSION

**Araneidae karyotypes under the light of phylogenetic and taxonomic studies.**—The karyotype configuration  $2n♂ = 22 + X_1X_2$  with acro/telocentric chromosomes found in the majority of species analyzed here corresponds to the most frequent complement in Araneidae (Table 2) (Araujo et al. 2021). Even though only 1.9% of all species of this family has been cytogenetically studied, the basic karyotype features seem to be relatively homogeneous in this group, with occasional rearrangements in some internal clades (Araujo et al. 2021; World Spider Catalog 2021).

“*Micrathenines*”: following Scharff et al. (2020), this clade clusters the genera *Acacesia* Simon, 1895, *Scoloderus* Simon, 1887, *Ocrepeira* Marx, 1883, *Verrucosa* McCook, 1888 and *Micrathena* Sundevall, 1833, all of them occurring in the New World (World Spider Catalog 2021). Within this clade are two groups, one clustering *Acacesia*, *Ocrepeira* and *Scoloderus* and another containing *Micrathena* and *Verrucosa* (see fig. 3 in Scharff et al. 2020).

Here, we show the first chromosomal data for *Micrathenines* with members of both *Micrathenine* clades. *Acacesia benigna* presents the standard karyotype of the family, while *Verrucosa meridionalis* and *V. scapofracta* have a much larger diploid number and unusual SCS (Table 2) (Fig. 4). Thus, the events which generated the chromosomal patterns in *Verrucosa* probably are restricted to the group comprising *Micrathena* and *Verrucosa* or occurred only in *Verrucosa* species. Therefore, karyological studies in *Micrathena* are needed in a next step of the research. Independently, due to the differences of diploid number and SCS between *V. meridionalis* and *V. scapofracta* we could affirm that some rearrangements have already occurred after *Verrucosa* divergence (Table 2).

“*Eriophorines*”: A clade that includes species of the genera *Acanthepeira* Marx, 1883, *Alpaida* O. Pickard-Cambridge, 1889, *Eriophora* Simon, 1864, *Parawixia* F. O. Pickard-Cambridge, 1904 and *Wagneriana* F. O. Pickard-Cambridge, 1904 (Scharff et al. 2020). Only six species of this clade were cytogenetically analyzed up to now, belonging to the genera

*Alpaida*, *Parawixia* and *Wagneriana* (Fig. 4). *Alpaida leucogramma* (White, 1841) and *Parawixia velutina* (Taczanowski, 1878) showed the common  $2n\delta = 24 = 22 + X_1X_2$  (Araujo et al. 2011), the same chromosome configuration of both *A. bicornuta* and *P. bistrata* (present study), whilst *Alpaida truncata* (Keyserling, 1865), *Alpaida veniliae* (Keyserling, 1865), *Parawixia kochi* (Taczanowski, 1873) and *Wagneriana* sp. exhibit a reduction of one autosomal pair, with  $2n\delta = 22 = 20 + X_1X_2$  (Araujo et al. 2011).

The only species of *Alpaida* with mitotic metaphases analyzed were *A. leucogramma* (Araujo et al. 2011) and *A. bicornuta* (present study). Despite both species presenting  $2n\delta = 24 = 22 + X_1X_2$ , the size of the largest autosomal pair of *A. bicornuta* corresponds to almost twice the size of the second pair (Fig. 1C), a chromosomal characteristic that was not observed in *A. leucogramma*. Thus, *A. bicornuta* possesses a complement different from the rest of the *Alpaida* species that have been cytogenetically studied to date.

*Parawixia bistrata* and *P. velutina* possess the same complement (present study; Araujo et al. 2011), suggesting a closer relationship chromosomal between them than to *P. kochi*. Unfortunately, there is no phylogeny of the genus so far that corroborates any hypothesis related to the species of *Parawixia*. However, the few *Parawixia* species included in phylogenetic analysis of the Araneidae by Scharff et al. (2020) showed as polyphyletic (part belonging to “Eriophorines” and part to “Backbourkiines”). Molecular data have been used to find species-specific characters, but are relatively scarce in Araneae (Agnarsson et al. 2013; Garrison et al. 2016; Scharff et al. 2020), and morphological characters have limitations for determining species in some groups (Scharff et al. 2020). Better approaches to these cases are those that include characteristics of different natures, such as behavior data (Coddington 1986, 1990; Dimitrov et al. 2017) and cytogenetic data (Araujo et al. 2008, 2011; Řezáč et al. 2018), for example. The karyotype of some *Parawixia* species presented variability and can be helpful in further studies of this polyphyletic genus, as suggested by Araujo et al. (2011). Cytogenetic studies in all “Eriophorines” genera are needed to establish a better understanding of the chromosome evolution in the clade.

“*Gasteracanthines*”: despite this clade possessing nine genera, the vast majority of species richness belongs to *Gasteracantha* Sundevall, 1833 (Scharff et al. 2020; World Spider Catalog 2021). Cytogenetically, only *Macracantha hasselti* (C. L. Koch, 1837) and *Gasteracantha kuhli* C. L. Koch, 1837 were analyzed and found to exhibit  $2n\delta = 16 = 14 + X_1X_2$  (Datta & Chatterjee 1983, 1988), diverging from diploid number  $2n\delta = 24 = 22 + X_1X_2$  of *G. cancriformis* studied here (Fig. 4). In the Araneidae, several instances of intrageneric reduction in diploid number have been reported. In *Larinioides* Caporiacco, 1934 these range from  $2n\delta = 14$  in *Larinioides patagiatus* (Clerck, 1757) to  $2n\delta = 23$  and  $24$  in *L. cornutus* (Clerck, 1757) (Berry 1906; Painter 1914; Hackman 1948; Qingtao et al. 1998, 1999). In *Neoscona* Simon, 1864 diploid numbers of  $2n\delta = 13, 14, 21, 23$  and  $24$  have been reported (Suzuki 1951a, b; Mittal 1960, 1966a; Datta & Chatterjee 1983, 1988; Parida & Sharma 1987; Sharma & Parida 1987; Amalin 1988; Amalin et al. 1993; Doan & Paliulis 2009; Prakash & Prakash 2014). Finally, *Alpaida* and *Para-*

*wixia* have just the reduction of one autosomal pair (Araujo et al. 2011).

A recent work that used morphology and DNA sequence data in a review of Gasteracanthines transferred *G. hasselti* to the genus *Macracantha* (Macharoenboon et al. 2021) corroborating the study of Tan et al. (2019), who found a paraphyly in *Gasteracantha*, specifically related to *G. hasselti*. However, both analyses used limited taxon sampling for a robust phylogeny of Gasteracanthines, and Macharoenboon et al. (2021) comment that the taxonomy in the group is complicated due to the difficulty of finding male specimens for morphological and molecular studies. The karyotype data suggest a closer proximity between *M. hasselti* and *G. kuhli* than between *G. cancriformis* and *G. kuhli*, which is concordant with the geographic distribution of these three species (Datta & Chatterjee 1983, 1988; World Spider Catalog 2021). Thus, further cytogenetic, morphological and phylogeographic work with several species are needed to help clarifying the relationships in this group.

*Actinosoma pentacanthum* and *Dubiepeira* sp.: the two largest phylogenies of the family do not include *Actinosoma* Holmberg, 1883 and *Dubiepeira* Levi, 1991 (Scharff & Coddington 1997; Scharff et al. 2020). However, the phylogenetic inference of *Wagneriana* by Cabra-García & Hormiga (2020) suggests a proximity of *Actinosoma* with *Alpaida*, *Rubrepeira* Levi, 1992 and some species of *Wagneriana*. From the cytogenetic point of view, *A. pentacanthum* presents the same diploid number ( $2n\delta = 24$ ) found in two of the four *Alpaida* species chromosomally analyzed up to now (*A. leucogramma* and *A. benigna*), whilst the other two *Alpaida* species and *Wagneriana* sp. presents  $2n\delta = 22$  (present study; Araujo et al. 2011), though further studies in these related genera are needed to determine more accurately the relationship among them.

In spite of the fact that *Dubiepeira* was not sampled in any phylogeny, Levi (1991) notice that females of *Dubiepeira* and *Araneus* Clerck, 1757 are very similar. Interestingly, *A. ventricosus* presented one of the highest diploid numbers in the Araneidae ( $2n\delta = 32, 46$  and  $49$ ) (Suzuki 1951a, b; Zhang & Tong 1990; Youju et al. 1993), a characteristic also observed in *Dubiepeira* sp. ( $2n\delta = 41$ ) in the present study and the same SCS ( $\delta X_1X_2X_3/\delta X_1X_1X_2X_2X_3X_3$ ) in one analyzed population (Youju et al. 1993). Moreover, *Araneus* is well known in the literature as a polyphyletic genus in which many unresolved species are inserted (Scharff & Coddington 1997; Scharff et al. 2020), thus, the karyotype found in *Dubiepeira* sp. suggests a proximity with *A. ventricosus* and chromosomal characters may be an alternative to help clarify relationships within the polyphyletic genus *Araneus*.

**Chromosomal evolution of the araneid karyotypes with high diploid numbers.**—Based on the high diploid numbers found in Mesothelae and several clades of Mygalomorphae, the chromosomal evolution in Araneae has been suggested to occur mainly through rearrangements that decrease the diploid number, such as Robertsonian fusions (Suzuki 1954; Král et al. 2006; Araujo et al. 2021). Nevertheless, even in the clades that possess mostly species with relatively low diploid numbers—such as the Araneidae—chromosomal fissions may occur, originating karyotypes with high diploid numbers like those found in *Dubiepeira* sp., *V. meridionalis* and *V.*

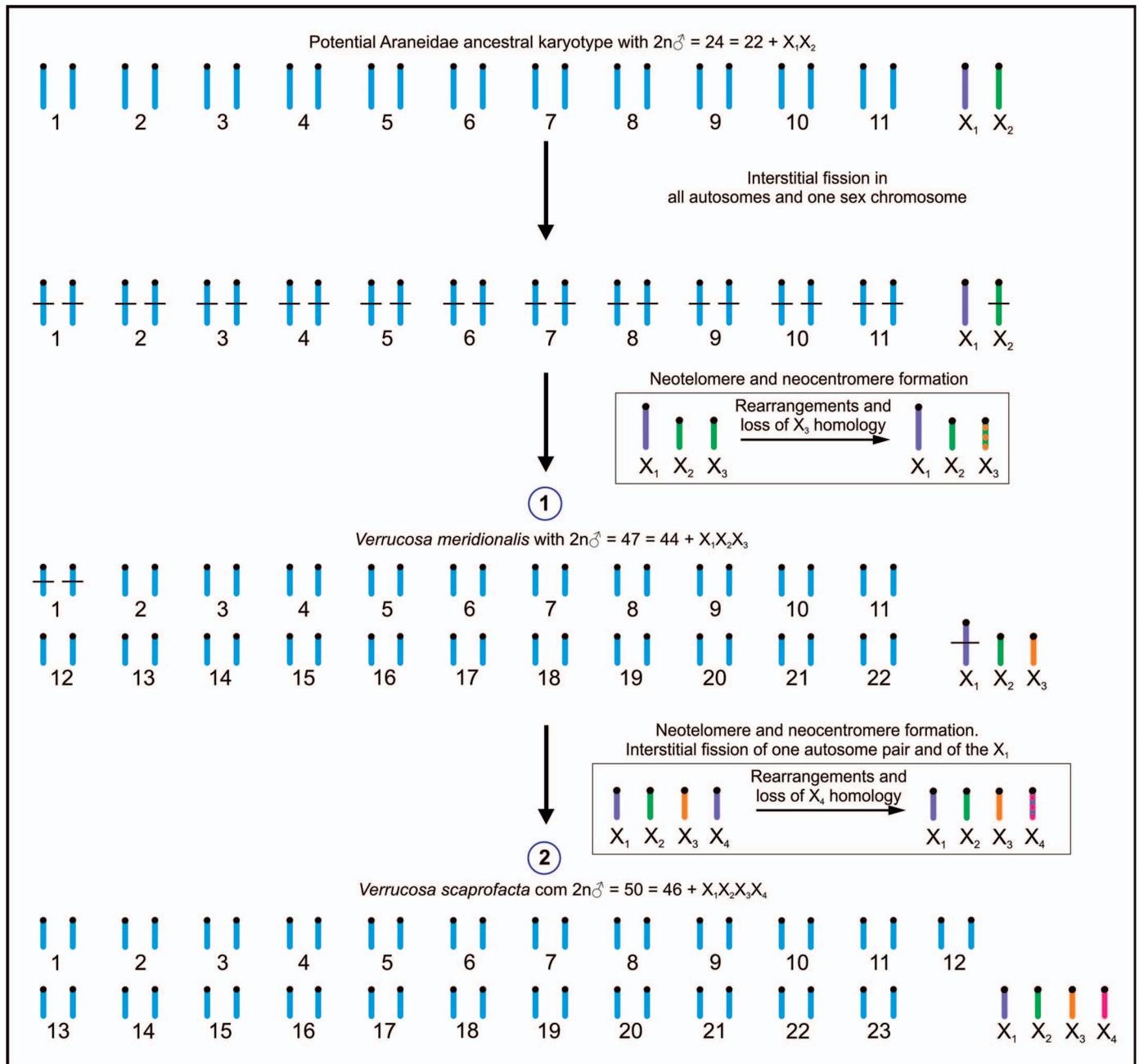


Figure 5.—Scheme showing hypothetical origins of the karyotypes  $2n\delta = 47 = 44 + X_1X_2X_3$  and  $2n\delta = 50 = 46 + X_1X_2X_3X_4$ , found in *Verrucosa meridionalis* and *Verrucosa scapofracta*, respectively. The rearrangement events represented by the numbers “1” and “2” would have occurred within the *Verrucosa* genus, as represented by the same numbers on “Micrathenines” (detail in Fig. 4).

*scapofracta* analyzed here. Interestingly, the  $2n\delta = 50$  of the latter species is the largest known diploid number within all Entelegynae spiders. A former study on *Agelenopsis naevia* (Walckenaer, 1841) (Agelenidae) mentioned around 50 chromosomes, but emphasizes that chromosome counting is not accurate, which is understandable considering that the study relies on histological sections. The same author had already described around 40 chromosomes previously to the same species (Wallace 1905, 1909).

The SCS of the type  $\delta X_1X_2X_3/\text{♀} X_1X_1X_2X_2X_3X_3$  is commonly found in spider families such as Sparassidae

(Hackman 1948; Suzuki & Okada 1950; Bole-Gowda 1952; Suzuki 1952; Mittal 1961, 1966b; Datta & Chatterjee 1983; Rowell 1985, 1988, 1990, 1991; Parida & Sharma 1986, 1987; Srivastava & Shukla 1986; Sharma & Parida 1987; Hancock & Rowell 1995), Agelenidae, in the genus *Tegenaria* Latreille, 1804 and Tetragnathidae, in the genus *Leucauge* White, 1841 (Sokólska 1925; Sharma et al. 1959; Datta & Chatterjee 1983, 1988; Xiuzhen et al. 1996; Král 2007; Kořínková & Král 2013). However, in the Araneidae it is very rare, restricted to *Araneus ventricosus* (Youju et al. 1993), *Dubiepeira* sp. and *V. meridionalis* (present study).

The SCS of the type  $\delta X_1X_2X_3X_4/\delta X_1X_1X_2X_2X_3X_3X_4X_4$  found in *V. scapofracta*, has been observed in nine species of spiders so far (Datta & Chatterjee 1983, 1988; Král et al. 2011, 2013; Kořínková & Král 2013; Souza et al. 2021). Souza et al. (2021) suggest its independent appearance sometimes in the Araneae because of the phylogenetic distance between the clades that have this SCS.

Based on the chromosome characteristics above described, we hypothesize a scenario to the origins of the *Verrucosa* karyotypes, that we will start from a karyotype with  $2n\delta = 24$ ,  $X_1X_2$ , because it is also present in other Araneoidea and probably the ancestor of the family (Araujo et al. 2021), which suffered events of chromosomal fission in all autosomal pairs and the  $X_2$  chromosome, originating a karyotype with  $2n\delta = 47$ ,  $X_1X_2X_3$ , as observed in *V. meridionalis*. In a next step, one autosomal pair and the  $X_1$  would be fissioned, originating the karyotype with  $2n\delta = 50$ ,  $X_1X_2X_3X_4$ , presented in *V. scapofracta* (Fig. 5).

Rearrangement events involving all chromosomes of the complement are called “all or nothing” and have already been described in the spider families Oxyopidae, Pholcidae, Sparassidae and Theridiidae (Rowell 1990; Stávale et al. 2010, 2011; Lomazi et al. 2018). However, more extensive research within the speciose genera *Verrucosa* and *Micrathena* can reveal karyotypes with intermediate diploid numbers, indicating a more gradual occurrence of the rearrangements.

Alternatively, the origin of the  $\delta X_1X_2X_3/\delta X_1X_1X_2X_2X_3X_3$  and  $\delta X_1X_2X_3X_4/\delta X_1X_1X_2X_2X_3X_3X_4X_4$  SCS could have happened through nondisjunctions followed by loss of homology of the neo X chromosomes, as suggested by some authors in other spider species (Postiglioni & Brum-Zorrilla 1981; Datta & Chatterjee 1988; Král et al. 2011; Araujo et al. 2012).

#### ACKNOWLEDGMENTS

This research was supported by Coordenacao de Aperfeiçoamento de Pessoal de Nivel Superior (CAPES)—Finance Code 001, Universidade Federal de Mato Grosso do Sul, Conselho Nacional de Pesquisa e Desenvolvimento Tecnológico (CNPq-PQ, ABD, grant #303903/2019-8) and Fundacao de Amparo a Pesquisa do Estado de Sao Paulo (FAPESP) process 99/05446-8. This study is part of the BIOTA/FAPESP—The Biodiversity Virtual Institute Program (www.biota.org.br).

#### LITERATURE CITED

- Agnarsson I, Coddington JA, Kuntner M. 2013. Systematics—progress in the study of spider diversity and evolution. Pp. 58–111. *In* Spider Research in the 21st Century: Trends and Perspectives. (Penney D, ed.). Manchester, Siri Scientific Press.
- Amalin DM. 1988. Morphology, life history and cytology of three orb-weaving spiders (Araneae: Araneidae) causing araneidism in the Philippines. *College, Laguna (Philippines)*, 1–79.
- Amalin DM, Barrion AA, Jayoma M. 1993. Comparative karyomorphology of two *Neoscona* species (Araneae: Araneidae). *Philippine Entomologist* 9:1–6.
- Araujo D, Mattos VF, Giroti AM, Kraeski MG, Carvalho LS, Brescovit AD. 2011. Cytogenetical characterization of six orb-weaver species and review of cytogenetical data for Araneidae. *Journal of Arachnology* 39:337–344. doi: 10.1636/CB10-88.1
- Araujo D, Rheims CA, Brescovit AD, Cella DM. 2008. Extreme degree of chromosome number variability in species of the spider genus *Scytodes* (Araneae, Haplogynae, Scytodidae). *Journal of Zoological Systematics and Evolutionary Research* 46:89–95. doi: 10.1111/j.1439-0469.2007.00457.x
- Araujo D, Schneider MC, Paula-Neto E, Cella DM. 2012. Sex chromosomes and meiosis in spiders: a review. Pp. 87–108. *In* Meiosis — Molecular Mechanisms and Cytogenetic Diversity. (Swan, A. ed.). Intech, Rijeka.
- Araujo D, Schneider MC, Paula-Neto E, Cella DM. 2021. The spider cytogenetic database. Version 9.5 Online at <http://www.arthropodcytogenetics.bio.br/spiderdatabase>, accessed on {April 2021}.
- Berry EH. 1906. The “Accessory chromosome” in *Epeira*. *Biological Bulletin* 11:193–201.
- Bole-Gowda BN. 1952. Studies on the chromosomes and the sex-determining mechanism in four hunting spiders (Sparassidae). *Proceedings of the Zoological Society of Bengal* 5:51–70.
- Cabra-García J, Hormiga G. 2020. Exploring the impact of morphology, multiple sequence alignment and choice of optimality criteria in phylogenetic inference: a case study with the Neotropical orb-weaving spider genus *Wagneriana* (Araneae: Araneidae). *Zoological Journal of the Linnean Society* 188:976–1151. doi: 10.1093/zoolin/zz088
- Coddington JA. 1986. The monophyletic origin of the orb web. Pp. 319–363. *In* Spiders: Webs, Behavior and Evolution. (Shear, W.A. ed.). Stanford University Press, Stanford, CA, USA.
- Coddington JA. 1990. Cladistics and spider classification: araneomorph phylogeny and the monophyly of orbweavers (Araneae: Araneomorphae; Orbiculariae). *Acta Zoologica Fennica* 190:75–87.
- Datta SN, Chatterjee K. 1983. Chromosome number and sex-determining system in fifty-two species of spiders from North-East India. *Chromosome Information Service* 35:6–8.
- Datta SN, Chatterjee K. 1988. Chromosomes and sex determination in 13 araneid spiders of North-Eastern India. *Genetica* 76:91–99. doi: 10.1007/BF00058807
- Dimitrov D, Benavides LR, Arnedo MA, Giribet G, Griswold CE, Scharff N, et al. 2017. Rounding up the usual suspects: a standard target-gene approach for resolving the interfamilial phylogenetic relationships of cribellate orb-weaving spiders with a new family-rank classification (Araneae, Araneoidea). *Cladistics* 33:221–250. doi: 10.1111/cla.12165
- Doan RN, Paliulis LV. 2009. Micromanipulation reveals an X0-XX sex determining system in the orb-weaving spider *Neoscona arabesca* (Walckenaer). *Hereditas* 146:180–182. doi: 10.1111/j.1601-5223.2009.02123.x
- Garrison NL, Rodriguez J, Agnarsson I, Coddington JA, Griswold CE, Hamilton CA, et al. 2016. Spider phylogenomics: untangling the Spider Tree of Life. *PeerJ* 4:e1719. doi: 10.7717/peerj.1719
- Green DM, Sessions SK. 1991. Amphibian cytogenetics and evolution. Pp. 431–432. *In* Appendix I, Nomenclature for Chromosomes. Academic Press.
- Hackman W. 1948. Chromosomen studien an Araneen mit besonderer berücksichtigung der geschlechtschromosomen. *Acta Zoologica Fennica* 54:1–101.
- Hancock AJ, Rowell DM. 1995. A chromosomal hybrid zone in the Australian huntsman spider, *Delena cancerides* (Araneae: Sparassidae). Evidence for a hybrid zone near Canberra, Australia. *Australian Journal of Zoology* 43:173–180. doi: 10.1071/ZO9950173
- Kořínková T, Král J. 2013. Karyotypes, sex chromosomes, and meiotic division. Pp. 159–171. *In* Spider Ecophysiology. (Nentwig W. ed.). Springer-Verlag, Berlin.
- Král J. 2007. Evolution of multiple sex chromosomes in the spider genus *Malthonica* (Araneae: Agelenidae) indicates unique structure

- of the spider sex chromosome systems. *Chromosome Research* 15:863–879.
- Král J, Kořínková T, Forman M, Krkavcová L. 2011. Insights into the meiotic behavior and evolution of multiple sex chromosome systems in spiders. *Cytogenetic and Genome Research* 133:43–66. doi: 10.1159/000323497
- Král J, Kořínková T, Krkavcová L, Musilová J, Forman M, Herrera IMA, et al. 2013. Evolution of karyotype, sex chromosomes, and meiosis in mygalomorph spiders (Araneae: Mygalomorphae). *Biological Journal of the Linnean Society* 109:377–408. doi: 10.1111/bij.12056
- Král J, Musilová J, Štěhlavský F, Řezáč M, Akan Z, Edwards RL, et al. 2006. Evolution of the karyotype and sex chromosome systems in basal clades of araneomorph spiders (Araneae: Araneomorphae). *Chromosome Research* 14:859–880. doi: 10.1007/s10577-006-1095-9
- Levan AK, Fredga K, Sandberg AA. 1964. Nomenclature for centromeric position on chromosomes. *Hereditas* 52:201–220. doi: 10.1111/j.1601-5223.1964.tb01953.x
- Levi HW. 1991. The Neotropical and Mexican species of the orb-weaver genera *Araneus*, *Dubiepeira*, and *Aculepeira* (Araneae: Araneidae). *Bulletin of the Museum of Comparative Zoology* 152:167–315.
- Lomazi RC, Araujo D, Carvalho LS, Schneider MC. 2018. Small pholcids (Araneae: Synspermiata) with big surprises: the lowest diploid number in spiders with monocentric chromosomes. *Journal of Arachnology* 46:45–49.
- Macharoenboon K, Siritwut W, Jeratthitikul E. 2021. A review of the taxonomy of spiny-backed orb-weaving spiders of the subfamily Gasteracanthinae (Araneae, Araneidae) in Thailand. *Zookeys* 1032:17–62.
- Mittal OP. 1960. Chromosome number and sex mechanism in twenty species of the Indian spiders. *Research Bulletin (N.S.) of the Panjab University* 11:245–247.
- Mittal OP. 1961. Chromosome number and sex mechanism in twenty-one species of the Indian spiders. *Research Bulletin (N.S.) of the Panjab University* 12:271–273.
- Mittal OP. 1966a. Karyological studies on the Indian spiders VI. Chromosome number and sex-determining mechanism in the family Araneidae. *Research Bulletin (N.S.) of the Panjab University* 17:335–351.
- Mittal OP. 1966b. Karyological studies on Indian spiders IV. Chromosomes in relation to taxonomy in Eusparassidae, Selenopidae and Thomosidae. *Genetica* 37:205–234.
- Painter TS. 1914. Spermatogenesis in spiders. *Zoologische Jahrbucher Abteilung fuer Anatomie und Ontogenie der Tiere* 38:509–576.
- Parida BB, Sharma NN. 1986. Karyotype and spermatogenesis in an Indian hunting spider, *Sparassus* sp. (Sparassidae: Arachnida) with multiple sex chromosomes. *Chromosome Information Service* 40:28–30.
- Parida BB, Sharma NN. 1987. Chromosome number, sex mechanism and genome size in 27 species of Indian spiders. *Chromosome Information Service* 43:11–13.
- Postiglioni A, Brum-Zorrilla N. 1981. Karyological studies on Uruguayan spiders I. Sex chromosomes in spiders of the genus *Lycosa* (Araneae-Lycosidae). *Genetica* 56:47–53.
- Prakash A, Prakash S. 2014. Cytogenetical investigations on spiders of semi-arid areas. *Indian Journal of Arachnology* 3:40–54.
- Qingtao Z, Hong H, Bing Y, Fengxiang L, Jian C. 1998. An observation of the spider chromosomes by single embryo-cell squash technique. *Acta Arachnologica Sinica* 7:39–41.
- Qingtao Z, Bing Y, Hong H, Jian C, Feng-Xiang L. 1999. On karyotype polymorphism in *Lariniodes cornuta*. *Acta Zoologica Sinica* 45:238–240.
- Rasband WS. 1997–2020. ImageJ, U. S. National Institutes of Health, Bethesda, Maryland, USA. Online at <http://imagej.nih.gov/ij/>
- Řezáč M, Arnedo MA, Opatova V, Musilová J, Řezáčová V, Král J. 2018. Taxonomic revision and insights into the speciation mode of the spider *Dysdera erythrina* species-complex (Araneae: Dysderidae): sibling species with sympatric distributions. *Invertebrate Systematics* 32:10–54. doi: 10.1071/IS16071
- Rowell DM. 1985. Complex sex-linked fusion heterozygosity in the Australian huntsman spider *Delena cancerides* (Araneae: Sparassidae). *Chromosoma* 93:169–176. doi: 10.1007/BF00293165
- Rowell DM. 1988. The chromosomal constitution of *Delena cancerides* Walck. (Araneae: Sparassidae) and its role in the maintenance of social behaviour. *The Australian Entomological Society Miscellaneous Publication* 5:107–111.
- Rowell DM. 1990. Fixed fusion heterozygosity in *Delena cancerides* Walck. (Araneae: Sparassidae): an alternative to speciation by monobrachial fusion. *Genetica* 80:139–157. doi: 10.1007/BF00127134
- Rowell DM. 1991. Chromosomal fusion and meiotic behaviour in *Delena cancerides* (Araneae: Sparassidae). I. Chromosome pairing and X-chromosome segregation. *Genome* 34:561–566. doi: 10.1139/g91-086
- Sakamoto Y, Zacaro AA. 2009. LEVAN, an ImageJ plugin for morphological cytogenetic analysis of mitotic and meiotic chromosomes. Initial version. Plugin JAVA of opencode accessible in <http://rsbweb.nih.gov/ij/>
- Scharff N, Coddington JA. 1997. A phylogenetic analysis of the orb-weaving spider family Araneidae (Arachnida, Araneae). *Zoological Journal of the Linnean Society* 120:355–434. doi: 10.1111/j.1096-3642.1997.tb01281.x
- Scharff N, Coddington JA, Blackledge TA, Agnarsson I, Framenau VW, Szűts T et al. 2020. Phylogeny of the orb-weaving spider family Araneidae (Araneae: Araneoidea). *Cladistics* 36:1–21. doi: 10.1111/cla.12382
- Sharma GP, Jande SS, Tandon KK. 1959. Cytological studies on the Indian spiders IV. Chromosome complement and meiosis in *Selenops radiatus* Latreille (Selenopidae) and *Leucauge decorata* (Blackwall) (Tetragnathidae) with special reference to XXX0-type of male sex determining mechanism. *Research Bulletin (N.S.) of the Panjab University* 10:73–80.
- Sharma NN, Parida BB. 1987. Study of chromosomes in spiders from Orissa. *Pranikee* 8:71–76.
- Sokólska J. 1925. Les hétérochromosomes pendant la spermatogénèse de l'Araignée domestique (*Tegenaria domestica* Cl.). *Bulletin de l'Acad. Polonaise des Sciences* 3B:477–491.
- Souza LHB, Silva BC, Costa CC, Brescovit AD, Rincão MP, Dias AL et al. 2021. First chromosomal analysis in Deinopidae (Araneae) reveals Sex Chromosome System  $X_1X_2X_3X_4$ , B chromosomes and polymorphism for centric fusion. *Zoology* 146:125906. doi: 10.1016/j.zool.2021.125906
- Srivastava MDL, Shukla S. 1986. Chromosome number and sex-determining mechanism in forty-seven species of Indian spiders. *Chromosome Information Service* 41:23–26.
- Stávale LM, Schneider MC, Araujo D, Brescovit AD, Cella DM. 2010. Chromosomes of Theridiidae spiders (Entelegynae): interspecific karyotype diversity in *Argyrodes* and diploid number intraspecific variability in *Nesticodes rufipes*. *Genetics and Molecular Biology* 33:663–668.
- Stávale LM, Schneider MC, Brescovit AD, Cella DM. 2011. Chromosomal characteristics and karyotype evolution of Oxyopidae spiders (Araneae, Entelegynae). *Genetics and Molecular Research* 10:752–763.
- Suzuki S. 1951a. Karyotypes in two families of spiders, Salticidae and Argiopidae. *Zoological Magazine* 60:3–4.
- Suzuki S. 1951b. Cytological studies in spiders. I. A comparative study of the chromosomes in the family Argiopidae. *Journal of Science of the Hiroshima University. Series B. Division 1* 12:67–98.
- Suzuki S. 1952. Cytological studies in spiders II. Chromosomal

- investigation in twenty two species of spiders belonging to the four families, Clubionidae, Sparassidae, Thomisidae and Oxyopidae, which constitute Clubionoidea, with special reference to sex chromosomes. *Journal of Science of the Hiroshima University. Series B. Division I* 13:1–52.
- Suzuki S. 1954. Cytological studies in spiders. III. Studies on the chromosomes of fifty-seven species of spiders belonging to seventeen families, with general considerations on chromosomal evolution. *Journal of science of the Hiroshima University. Series B. Division I* 15:23–136.
- Suzuki S, Okada A. 1950. A study on the chromosomes of a spider, *Heteropoda venatoria*, with special reference to X1-, X2- and X3-chromosomes. *Journal of Science of the Hiroshima University. Series B. Division I* 11:29–44.
- Tan J, Chan ZY, Ong CA, Yong HS. 2019. Phylogenetic relationships of *Actinacantha* Simon, *Gasteracantha* Sundevall, *Macracantha* Hasselt and *Thelacantha* Simon spiny orbweavers (Araneae: Araneidae) in Peninsular Malaysia. *Raffles Bulletin of Zoology* 67:32–55. doi: 10.26107/RBZ-2019-0003
- Xiuzhen W, Youju W, Zhenling Y, Sujuan C, Ning L. 1996. On chromosomes of the *Tegenaria domestica* (Araneida: Agelenidae). *Acta Arachnologica Sinica* 5:141–144.
- Wallace LB. 1905. The spermatogenesis of the spider. *Biological Bulletin* 8:169–184.
- Wallace LB. 1909. The spermatogenesis of *Agalena naevia*. *Biological Bulletin* 17:120–161.
- World Spider Catalog. 2021. World Spider Catalog. Version 22 Natural History Museum, Bern. Online at <http://wsc.nmbe.ch>, accessed on {April 2021}. doi: 10.24436/2
- Youju W, Daxiang S, Xiuzhen W, Zhenling Y. 1993. Preliminary studies on the chromosome of four species of spiders. *Acta Arachnologica Sinica* 2:110–113.
- Zhang YJ, Tong SJ. 1990. The routine method for preparing the chromosomes in spiders. *Chinese Journal of Zoology* 25:30–31.
- Manuscript received 12 September 2020, revised 24 April 2021, accepted 9 August 2021.*